

Published on Web 09/13/2010

Total Synthesis of Chivosazole F

Tobias Brodmann,[†] Dominic Janssen,[‡] and Markus Kalesse^{*,†,§}

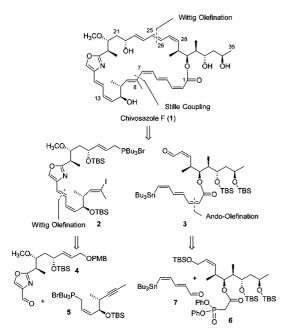
Centre for Biomolecular Drug Research (BMWZ), Leibniz Universität Hannover, Schneiderberg 1b, D-30167 Hannover, Germany, Centre for Cancer Research & Cell Biology, Queen's University Belfast, 97 Lisburn Road, Belfast BT9 7BL, U.K., and Helmholtz Centre for Infection Research, Inhoffenstrasse 7, D-38124 Braunschweig, Germany

Received August 13, 2010; E-mail: markus.kalesse@oci.uni-hannover.de

Abstract: The first synthesis of the highly biologically active chivosazole F is described. It features an intramolecular Stille coupling for the macrolactone formation and thereby circumvents the problem of isomerization associated with the tetraene segment. Additionally, the synthesis confirms the structure which has been proposed based solely on a combination of NMR/computational methods and genetic analysis.

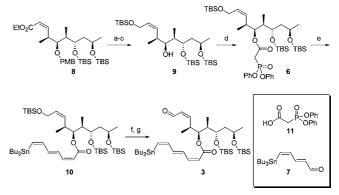
The family of the chivosazoles contains seven 31-membered macrolides which possess a remarkable biological profile.¹ They exhibit antifungal and cytotoxic activity against mouse fibroblasts (L-929) and HeLa cells (IC₅₀ 9 ng/mL for both cell lines)^{1a} and reduce actin polymerization.² In these studies it became apparent that the chivosazoles act on F-actin differently than proposed for cytochalasin D, chondramide, or rhizopodin.³ Structurally, chivosazole F differs from the other members of this family by lacking the 6-deoxyglucose derivative (chivose) at C11.

Scheme 1



At the outset of our investigations we proposed the unknown configurations of the 10 stereogenic centers through a combination of NOE experiments and computational methods.⁴ The configurations at the secondary alcohols were confirmed by genetic analysis put forward by Reid and Caffrey.4,5 Even with this state of knowledge total synthesis was required to confirm the configurations of the 10 stereocenters. As a consequence of the lability of polyenes, in particular when Z-configured double bonds are involved, we embarked on a strategy that established the two sensitive polyene segments in the last steps of the macrolactone formation. Additionally, the recently published synthesis on the southern hemisphere of chivosazole F by Paterson et al. confirmed the sensitive nature of the tetraenoate subunit.⁶ We proposed the final connections to be made between C25 and C26 by a Wittig olefination and between C7 and C8 by an intramolecular Stille coupling. During our studies on the synthesis of chivotriene we realized that the best selectivities were observed when segment 2 serves as the Wittig reagent and segment **3** as the corresponding aldehyde (Scheme 1).⁷ The assembly of segment 3 started from unsaturated ester 8.8 Removal of the PMB group, DiBAl-H reduction, and protection of the primary alcohol as the TBS ether was followed by esterification with phosphonate $11.^9$ Finally, olefination of 6 with aldehyde 7^{10} provided triene 10 in 80% yield as a 1:1 mixture of the Z and Eisomers (Scheme 2). Gratifyingly, the mixture of isomers could be separated by simple column chromatography and the desired olefin was used for the subsequent transformations.

Scheme 2^a



^a Reagents and conditions: (a) DDQ, CH₂Cl₂/H₂O, 70%; (b) DiBAl-H; (c) TBSCl, imidazole, 78% (over 2 steps); (d) 11, DMAP, Et₃N, 2,4,6trichlorobenzoyl chloride, toluene, 87%; (e) 7, NaH, THF, 80%, Z/E = 1:1; (f) HF-pyridine, pyridine, 51%; (g) MnO₂, CH₂Cl₂, 85%.

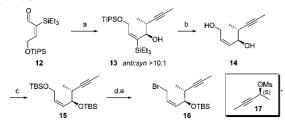
For the construction of the western hemisphere, we aimed to establish the anti relationship in segment 5 through a Marshall-Tamaru reaction. In order to circumvent the modest selectivities of the Marshall reaction in transformations with unsaturated aldehydes,11 we employed aldehyde 12 with a triethyl silyl group in the α -position to enhance the diastereoselectivity in this reaction.

Leibniz Universität Hannover.

[‡] Queen's University Belfast. [§] Helmholtz Centre for Infection Research.

Using the indium protocol developed by Marshall et al.^{11a} and S-configured propargylic mesylate 17, the reaction was performed with high selectivities (anti/syn > 10:1) and provided 13 in good yields (68%) (Scheme 3). The subsequent standard transformations established allyl bromide 16.

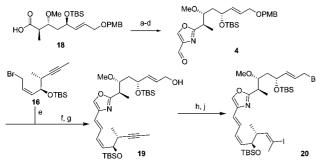
Scheme 3^e



^a Reagents and conditions: (a) 17, Pd(OAc)₂, •PPh₃, InI, THF-HMPA, 68%; (b) TBAF (5% H₂O), 83%; (c) TBSOTf, 2,6-lutidine, 99%; (d) PPTS, MeOH, 65%; (e) CBr₄, PPh₃, 99%.

Aldehyde 4 was derived from acid 18 through a sequence of esterification and standard oxazole formation.¹² DiBAl-H reduction established the aldehyde functionality required for the Wittig olefination with 5.¹³ The subsequent olefination installed the new double bond with high selectivity (E/Z > 10:1). At that stage it proved beneficial to remove the PMB protecting group prior to the functionalization of the alkyne. For this transformation the palladium-catalyzed hydrostannylation provided the highest yields and regioselectivities. Finally, the allylic alcohol was converted to the corresponding allyl bromide 20 (Scheme 4).

Scheme 4^e

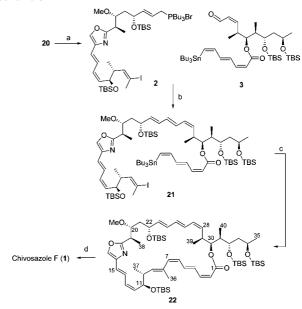


^a Reagents and conditions: (a) SerOMe•HCl, EDC•HCl, HOBT, *i*-Pr₂NEt, 78%; (b) DAST, CH₂Cl₂; (c) DBU, BrCCl₃, CH₂Cl₂, 0 °C; (d) DiBAl-H, toluene, 70% 3 steps; (e) PBu₃, MeCN; (f) KOtBu, toluene, 91% (over 2 steps); (g) DDQ, CH₂Cl₂/H₂O, 63%; (h) (i) Bu₃SnH, (PPh₃)₂PdCl₂; (ii) I₂, THF, 67%; (j) (i) MsCl, Et₃N; (ii) LiBr, THF, 82%.

With both hemispheres in hand we continued to establish the E-configured double bond between C25 and C26 using a Wittig reaction. Without further functional group transformations the intramolecular Stille¹⁴ reaction followed to establish the carbon framework of chivosazole. The final global deprotection using buffered HF then completed the total synthesis of chivosazole F (Scheme 5). The spectroscopic data (¹H NMR, ¹³C NMR, HRMS, optical rotation) were in all respect identical to the data originally reported by Höfle and co-workers thus confirming the previously made configurational assignment.

In summary, the first total synthesis of (-)-chivosazole F has been achieved through a strategy that establishes the delicate polyene systems using late-stage Wittig reactions and an intramolecular Stille coupling as the pivotal transformations for their constructions.

Acknowledgment. Generous financial support by the Deutsche Forschungsgemeinschaft (KA 913/14-1) and the Fonds der Che-



^a Reagents and conditions: (a) PBu₃, MeCN; (b) KOtBu, toluene, 48% (over 2 steps); (c) PdCl₂(PhCN)₂, DMF; (d) HF-pyridine, pyridine, 18% (over 2 steps).

mischen Industrie for a FONDS-fellowship for T.B. is gratefully acknowledged. We thank R. Jansen (HZI) for providing an authentic sample and S. Eichner for HPLC assistance.

Supporting Information Available: Spectroscopic and analytical data as well as experimental procedures for all compounds described herein. This material is available free of charge via the Internet at http:// pubs.acs.org.

References

Scheme 5^e

- (1) (a) Jansen, R.; Irschik, H.; Reichenbach, H.; Höfle, G. Liebigs Ann. Chem. **1997**, 8, 1725–1732. (b) Brodmann, T.; Lorenz, M.; Schäckel, R.; Simsek, S.; Kalesse, M. *Synlett* **2009**, *2*, 174–192.
- Diestel, R.; Irschik, H.; Jansen, R.; Khalil, M. W.; Reichenbach, H.; Sasse, F. ChemBioChem 2009, 10, 2900–2903. (2)
- For cytochalasin D, see: (a) Lin, D. C.; Lin, S. Proc. Natl. Acad. Sci. U.S.A. 1979, 76, 2345–2349. (b) Flanagan, M. D.; Lin, S. J. Biol. Chem. 1980, (3)255, 835–838. (c) MacLean-Fletcher, S.; Pollard, T. D. Cell **1980**, 20, 329– 341. For condramides, see: (d) Kunze, B.; Jansen, R.; Sasse, F.; Höfle, G.; Reichenbach, H. J. Antibiot. 1995, 48, 1262-1266. (e) Eggert, U.; Diestel, R.; Sasse, F.; Jansen, R.; Kunze, B.; Kalesse, M. Angew. Chem., Int. Ed. **2008**, 47, 6478–6482. For rizopodin, see: (f) Sasse, F.; Steinmetz, H.; Höfle, G.; Reichenbach, H. J. Antibiot. **1993**, 46, 741–748.
- (4) Janssen, D.; Albert, D.; Jansen, R.; Müller, R.; Kalesse, M. Angew. Chem., Int. Ed. 2007, 46, 4898-4901.
- (5) (a) Reid, R.; Piagentini, M.; Rodriguez, E.; Ashley, G.; Viswanathan, N.; Carney, J.; Santi, D. V.; Hutchinson, C. R.; McDaniel, R. *Biochemistry* 2003, 42, 72–79. (b) Caffrey, P. *ChemBioChem* 2003, 4, 654–657.
- (6) Paterson, I.; Kan, S. B. J.; Gibson, L. J. Org. Lett. 2010, 12, 3724–3727.
 (7) Brodmann, T.; Janssen, D.; Sasse, F.; Irschik, H.; Jansen, R.; Müller, R.; Kelasso, M. Fur, L. Org. Chem. 2010, 5155–5150.
- (7) Drommin, F., Janssen, D., Basser, F., Hsenn, H., Janssen, M. Eur. J. Org. Chem. 2010, 5155–5159.
 (8) Janssen, D.; Kalesse, M. Synlett 2007, 17, 2667–2670.
- (9) Ghosh, A. K.; Wang, Y.; Kim, J. T. J. Org. Chem. 2001, 66, 8973-8982.
- (10) Mapp, A. K.; Heathcock, C. H. J. Org. Chem. 1999, 64, 23–27.
- (11) (a) Marshall, J. A.; Eidam, P.; Schenk Eidam, H. J. Org. Chem. 2006, 71, 4840–4844. (b) Marshall, J. A. Chem. Rev. 2000, 100, 3163–3186.
- (12) (a) Williams, D. R.; Brooks, D. A.; Berliner, M. A. J. Am. Chem. Soc. 1999, 121, 4924–4925. (b) Phillips, A. J.; Uto, Y.; Wipf, P.; Reno, M. J.; Williams, D. R. Org. Lett. 2000, 2, 1165–1168.
- (13) (a) Tamura, R.; Kato, M.; Saegusa, K.; Kakihana, M.; Oda, D. J. Org. Chem. **1987**, 52, 4121–4124. (b) Tamura, R.; Saegusa, K.; Kakihana, M.; Oda, D. J. Org. Chem. **1988**, 53, 2723–2728.
- (a) Milstein, D.; Stille, J. K. J. Am. Chem. Soc. 1978, 100, 3636-3638. (14)For reviews, see: (b) Duncton, M. A. J.; Pattenden, G. J. Chem. Soc., Perkin Trans. 1 1999, 1235-1246. (c) Pattenden, G.; Sinclair, D. J. J. Organomet. Chem. 2002, 653, 261-268. (d) Nicolaou, K. C.; Bulger, P. G.; Sarlah, D. Angew. Chem., Int. Ed. 2005, 44, 4442-4489

JA107290S